Analogs of natural lipids. IV. Synthesis and properties of cyclopentanoid analogs of phosphatidic acid¹

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Abstract A new series of phosphatidic acid analogs has been synthesized in which the glycerol moiety of diacylglycerophosphoric acid is replaced by each of the three isomeric cyclopentane-1,2,3-triols (1,3/2, DL-1,2/3, and1,2,3/0). Of the seven possible configurational and positional phosphatidic acid analogs of this series, five isomers have been obtained and characterized by spectroscopic methods and microanalysis. Four of the five isomers are 1-(or 3-)phosphoryl derivatives, while the fifth is a 2-phosphate.

The analogs were prepared in configurationally pure form by unequivocal synthetic procedures involving selectively blocked intermediates; acyl migration was avoided by the use of mild deblocking procedures. The anhydrous lipid products, all of which are dipalmitoyl esters, are solids indefinitely stable at room temperature in the free acid or potassium salt form; they have chromatographic mobility and melting points similar to dipalmitoyl glycerophosphoric acid. The dipotassium salts bind water of hydration tenaciously, remaining hydrated after drying in vacuo at 100°C. NMR spectra of dimethyl esters of some of the analogs show nonequivalence of the two methyl groups, consistent with the diastereotopic nature of those groups.

In addition to their intrinsic interest as conformationally restricted acidic lipids, the analogs are now available as starting materials for the synthesis of the more complex acidic and amphoteric lipids required for our exploitation of these cyclopentanoid analogs as unique probes for the study of lipid-lipid and protein-lipid interactions.

Supplementary key words lipid analogs · conformation of phosphoglycerides · hydration of phosphatides · nuclear magnetic resonance spectroscopy · diastereotopic groups · cyclitols

This communication is part of a series that describes the systematic synthesis and study of a new class of analogs of natural lipids. Previously (2), we reported the synthesis of three homologous series of triglyceride analogs, derived from each of the three diastereoisomeric cyclopentane-1,2,3-triols 1, 2, and 3 (Fig. 1). This work has now been extended to include the synthesis of five (4-8) of the seven possible phosphatidic acid analogs (4-10) in which the glycerol moiety of naturally occuring PA is replaced by one or the other of the three cyclopentane-1,2,3-triols. The rationale for the program of synthesis we have undertaken is the expectation that the lipid analogs will provide an original probe into the conformational aspects of lipid-lipid and lipid-protein interaction in biological situations.

Because there is no covalent restriction of the rotation of the glycerol backbone of natural glycerides, it has been impossible hitherto to assess the contribution of the rotameric state of that backbone to lipid-lipid and lipid-protein interactions. On the other hand, the cyclopentanoid ring imposes severe restrictions, so that each of the analogs may correspond to only a small number of rotameric states of the backbone of corresponding glycerides and one may therefore hope to observe preferential behavior of one or more analogs in model systems for the interactions noted.

In addition, the analogs may be substrates for or inhibitors of lipid metabolizing enzymes, again sug-

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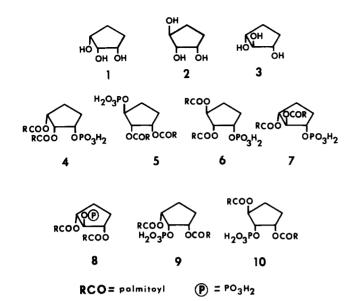
Abbreviations and nomenclature: CHCl₃, chloroform; d, doublet; 2,2-DMP, 2,2-dimethoxypropane; DPPC, diphenylphosphoryl chloride; EtOAc, ethyl acetate; EtOH, ethanol; Et₂O, diethyl ether; GLC, gas-liquid chromatography; HOAc, acetic acid; Ip, isopropylidene [(CH₃)₂C=]; m, multiplet; MeOH, methanol; NMR, nuclear magnetic resonance; PA, phosphatidic acid; PC, phosphatidylcholine; Pd-C, palladium-charcoal (10%); ppm, parts per million; s, singlet; TLC, thin-layer chromatography; TMS, tetramethylsilane; TROC, 2,2,2-trichloroethoxycarbonyl.

Cyclic compounds described in this paper are named according to the Tentative Rules for Nomenclature of Cyclitols (1). The names are derived from those of the parent cyclanes of which they are formal derivatives. A summary of these rules has been presented in an earlier communication (2).

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Fig. 1. Configuration of cyclopentanetriols and of cyclopentanoid analogs of phosphatidic acid. *1*: (1,2,3/0)-cyclopentane-1,2,3-triol; *2*: DL-(1,2/3)-cyclopentane-1,2,3-triol; *3*: (1,3/2)-cyclopentane-1,2,3-triol.

gesting the rotameric condition of the true substrates. Evidence supporting this contention has recently been obtained by Morrisett et al.⁴ who found marked differences in the physical properties of the five analogs 4-8 and also in their ability to bind to apolipoprotein-C-III from human serum. The conformational rationale of the analogs has been published (2). Finally, the PA analogs are now available as starting materials for the synthesis of more complex cyclopentanoid phospholipid analogs. The synthetic routes developed allow the preparation of gram quantities of each of the five compounds in an analytically and stereochemically pure state. In each case, however, the compound has been synthesized from DL- or mesostarting material, so that the unsymmetrical products (4-7) are DL mixtures while 8 is a meso compound. The synthesis of compounds 4-8 is described in this paper.

MATERIALS AND METHODS

Melting points were measured on a Kofler micro hot-stage (Arthur H. Thomas and Co., Philadelphia, PA) and are uncorrected. Refractive index was measured with an Abbé refractometer. Infrared spectra were measured with a Perkin-Elmer 237B spectrophotometer (Perkin-Elmer Corp., Norwalk, CT) and were calibrated with polystyrene. NMR spectra were recorded on Varian A-60 and A-60A spectrometers for dilute solutions (Varian Associates, Palo Alto, CA), with TMS as internal standard. Unless otherwise stated, the solvent used was ²H-chloroform. Chemical shifts are reported on the δ -scale in ppm downfield from TMS; chemical shifts are positive in this direction. Microanalyses were performed by Galbraith Laboratories, Knoxville, TN.

Analysis of reaction products by TLC and GLC was performed as described earlier (2); phosphates were visualized on TLC plates by spraying with the reagent of Dittmer and Lester (3).

Solutions of reaction products in organic solvents were dried with Na_2SO_4 before evaporation under reduced pressure on a rotary evaporator. Products of acylation and phosphorylation reactions were extracted into CHCl₃ and washed successively with water, ice-cold 1 N H₂SO₄, 5% NaHCO₃ and water.

Diphenylphosphorylchloride (DPPC) and 2,2,2trichloroethyl chloroformate were obtained from Aldrich Chemical Co. Inc., Milwaukee, WI. Palmitoyl chloride and palmitic anhydride were products of Nu-Chek Prep., Elysian, MN. Celite 545 was supplied by Fisher Scientific Co., Pittsburgh, PA, and was washed with chloroform-methanol 1:1 before use. Platinum oxide was obtained from Engelhard Industries Co., Newark, N. J. Acylation reactions were generally carried out in anhydrous pyridine as described earlier for triacyl derivatives (2); in one case (1,3/2-isomer, 26) the yield was marginally improved by using the fatty acid anhydride method of Cubero Robles and Van Den Berg (4). The dipalmitates were purified by silicic acid chromatography and recrystallization from methanol (2). O-Isopropylidene derivatives of diols were prepared as described previously (2). Epoxidations were carried out in widemouth brown bottles (2). Diphenyl esters of PA were hydrogenolyzed in quantitative yield over platinum in acetic acid. After centrifugation and evaporation of solvent, the PA analog was partitioned to remove cations according to a modification (5) of the procedure of Bligh and Dyer (6). Free PA was thus obtained as a sharply-melting white solid (Table 1). PA analogs (ca. 50 mg in CHCl₃, ca. 2 ml) were converted into potassium salts by titration to the phenolphthalein end point with KOH (0.2 N in MeOH-H₂O 98:2), and isolated in 90-95% yield as described by Kates et al. for phytanyl-ether PA analogs (5).

⁴ J. D. Morrisett, A. M. Gotto, K-Y Hong, L. C. Smith, A. J. Hancock, and H. Z. Sable. *Federation Proc.* **35**, 1506 (1976), Abstract No. 746. Presented at the 67th Annual Meeting, American Society of Biological Chemists, San Francisco, June 1976. A full manuscript is in preparation.

TABLE 1. Analytical data for analogs of phosphatidic acid and potassium phosphatidate

		Free Acids C ₃₇ H ₇₁ O ₈ P (674.9)			K₂ Salts, Trihydrates ^a C37H69O8PK2 · 3H2O (805.2)				K ₂ Salts, Dihydrates ^c C ₃₇ H ₆₉ O ₈ PK ₂ · 2H ₂ O (787.1)			
		C	Н	Р	C	Н	Р	K	K/P ^b	С	н	H_2O^d
Theory:		65.84	10.60	4.59	55.19	9.39	3.85	9.71	2.00	56.46	9.35	4.58
Compound	Configuration		Found			.	Found				Found	
4	1,2,3/0-(1P)	65.98	10.70	4.42	55.55	9.43	3.79	9.08	1.90	56.85	9.76	5.10 ^d
5	1,2/3-(3P)	65.91	10.70	4.45	55.50	9.60	3.78	9.40	1.97	56.41	9.55	
6	1,2/3-(1P)	65.94	10.51	4.49	55.00	9.05	3.71	9.17	1.96	56.50	9.75	
7	1,3/2-(1P)	66.08	10.55	4.36	55.51	9.85	3.53	9.40	2.11	56.70	9.91	
8	1,3/2-(2P)	66.29	10.27	4.38	55.10	9.65	3.66	9.25	2.00	55.65	9.51	

^a Compounds dried over P₂O₅ (12 hr at room temperature, 0.5 mm).

^b K/P is ratio (gram-atoms K)/(gram-atoms P).

^c Compounds dried over P_2O_5 (12 hr at room temperature, 0.5 mm), then dried further over P_2O_5 for 12 hr at 100°C (0.5 mm).

^d Analysis by Karl Fischer method.

Methylation of PA analogs

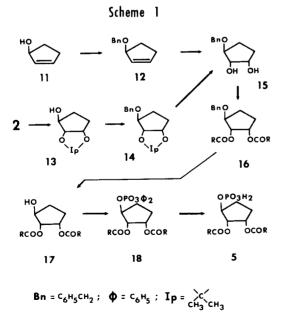
Each free PA analog was treated with excess diazomethane (7) to give the dimethyl ester. Evaporation gave the esters in quantitative yield. All five esters were sharply melting, gave one spot on TLC, and were stable on storage at room temperature, unlike related esters (8).

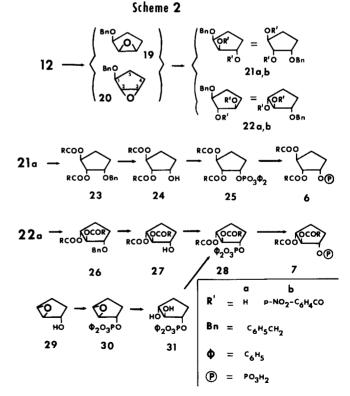
3). The acid stability of benzyl ethers allowed removal of the isopropylidene group (e.g., $14 \rightarrow 15$); the resulting diols were acylated directly either with palmitoyl chloride or with palmitic anhydride (4). The benzyl group was removed by hydrogenolysis over Pd, and the resulting unisomerized diglyceride analog was phosphorylated to give the diphenyl ester of the PA analog.

Two benzyl ethers (21a and 22a) were prepared by an indirect route. Epoxidation of O-benzyl-2-

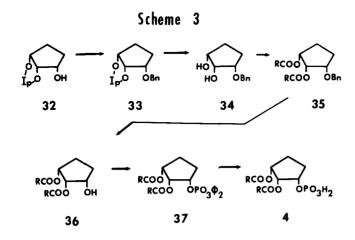
RESULTS AND DISCUSSION

Monobenzyl ethers of cyclopentanetriols were key intermediates in several of the synthetic sequences, i.e., $13 \rightarrow 14$ (Scheme 1); $32 \rightarrow 33$ (Scheme





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cyclopenten-1-ol 12 gave a mixture of the isomeric epoxides 19 and 20 in a ratio of 1:2. This selectivity is unexpectedly low (9, 10). Since acid hydrolysis of each of the epoxides gave both 21a and 22a, the mixture was not separated, but was hydrolyzed directly and the diols converted to the bis-O-p-nitrobenzoate esters; the latter were separated by fractional crystallization and the purified substances were saponified as needed. For study of the regioselectivity (11) of hydrolysis of 19 and 20 the isomers were separated by preparative TLC. Hydrolysis of 19 gave 21a and 22a in a ratio of 9:91, while 20 gave a ratio of 95:5. This selectivity is similar to that seen (11) in related systems.

The synthesis of ϑ , a symmetrical 2-phosphate, required a 1,3-blocked (1,3/2)-triol. The 1,3-benzylidene acetals of both glycerol (12) and *cis*-1,3-cyclopentanediol (13), as well as the 1,3-isobutylidene acetal⁵ are known. However, in this series the *p*nitrobenzylidene acetal $\vartheta \vartheta$ offered the advantages of both crystallinity and susceptibility to mild deblocking conditions. In this sequence, intramolecular migration is not possible since the diphenylphosphoryl group of $\vartheta \vartheta$ (Scheme 4) is *anti* to the regenerated hydroxyl groups.

DL-(1,2/3)-3-Phosphoryl analog

The benzyl diol 15 was synthesized by two routes (Scheme 1): one route required the triol acetonide 13 (14) and its benzyl derivative 14. A more direct route to 15 involved direct hydroxylation of the allylic benzyl ether 12, which is readily prepared from 11. Hydroxylation with alkaline KMnO₄ was highly selective, less than 1% of the (1,2,3/0) isomer being formed. All four benzyl diols (15, 21a, 22a, and 34) are thermally stable and can be distilled

in nitrogen without decomposition; however, they are prone to deterioration on storage in air at room temperature. Ideally, each diol was used in the next stage of the synthesis without delay, but the diols could be stored over nitrogen at -20° C, or as the *bis-p*-nitrobenzoates at room temperature.

Diol 15 was acylated as described for the synthesis of the *tris*-acylates (2). The *bis*-palmitate was a stable solid; hydrogenolysis over Pd-C gave the analog 17, which was phosphorylated to give 18. The latter was stored at 4°C since, for some monophenyl esters (15), phenol is liberated on prolonged storage at room temperature. Catalytic dephenylation of 18 with PtO₂ in acetic acid gave 5 smoothly, while PtO₂ in ethanol gave complex mixtures. The free acid form of 5 melted in the range reported for glycerophosphatidic acids (5, 16).

DL-(1,3/2)-1-phosphoryl analog

Two (1,3/2)-PA analogs were synthesized. The alltrans isomer 7 was synthesized by two independent routes (**Scheme 2**). The high regioselectivity (11) of hydrolysis of epoxide 30 gave pure 31, and the latter was acylated directly to give 28.

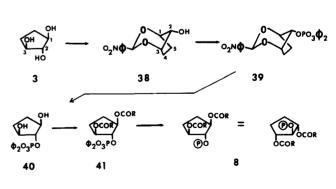
DL-(1,3/2)- and DL-(1,2/3)-1-phosphoryl analogs

A second route to 7 involves the intermediate benzyl ether 22a, which is a by-product in the formation of 1,2/3-isomer 21a (Scheme 2). The respective benzyl diols were acylated, debenzylated, and phosphorylated to give 28 and 25 in good yield. The phenyl groups were removed by hydrogenolysis over Pt.

DL-(1,2,3/0)-1-phosphoryl analog

Scheme 4

The all-*cis* analog 4 was synthesized by a route (Scheme 3) resembling that for analog 5. 1,2,3/0-Cyclopentanetriol acetonide 32 was obtained by reductive ring opening of DL-(1,2,3,4/0)-1,2-anhydro-



 $\phi = c_6 H_5$; $o_2 N \phi = p - N O_2 - c_6 H_4$; $\phi = P O_3 H_2$

⁵ N. Dennis and E. L. Eliel. Unpublished experiments. We thank Professor Eliel for providing this information.

cyclopentane-1,2,3,4-tetrol (2) and purified via its benzoate ester. Benzylation and hydrolysis gave 34, which was acylated and then converted into the pseudodiglyceride 36 without concomitant acyl group migration, as judged by TLC and melting behavior. However, prolonged exposure of 36 to unbuffered silicic acid did promote acyl migration, a property shared by the 1,2/3 isomer 24 (Scheme 2), as well known in glyceride chemistry (12).

2-O-Phosphoryl analog of PA

The symmetrical isomer ϑ , one of the three possible "abnormal" analogs having the phosphoryl moiety at position 2 of the ring (Fig. 1), has been synthesized via the *p*-nitrobenzylidene acetal ϑ . The synthetic sequence differed from that used for most of the other isomers, but resembled the route to the other all-*trans* isomer 7 (via ϑ 1) in that the diphenylphosphorylation step preceded all the others. Hydrogenolysis of $\vartheta \vartheta$ over Pd-C removed the acetal blocking group without affecting the phenyl groups (17, 18). Acylation followed by catalytic dephenylation gave ϑ .

Hydration of potassium salts

Elemental analyses showed that the free acid forms of 4-8 were anhydrous (Table 1). The equivalent weight of each isomer, determined by titration, agreed closely with the theoretical values. On the other hand, the elemental analysis showed that the vacuum-dried potassium salts exist as the trihydrates (Table 1). The water of hydration was particularly strongly bound: overnight drying at 100°C/0.5 mm removed only 1 mole of water. Confirmation was obtained by Karl Fischer analysis of the salt of the all-cis isomer 4 (Table 1). The tenacity of the hydration may be related to other observations on amphoteric lipids. Chapman and Fluck (19) related the temperature at which the lipid formed smectic mesophases in water to the major peaks observed in differential thermal analysis of the dry compounds. In effect this relates the degree of liquidity of the hydrocarbon side chains to the ease of disruption of phospholipid crystals by water. Williams and Chapman (20) showed that, for each molecule of PC, 10 molecules of water do not freeze at 0°C, and Bangham, deGier, and Greville (21) found that about 35 molecules of water, per molecule of PC, are osmotically inactive.

Melting point behavior of the PA analogs

The PA analogs melt in the range $73-84^{\circ}$ C (**Table 2**), while dipalmitoyl glycerophosphoric acid acid and its distearyl ether analog melt at $70-71^{\circ}$ C (16) and 69° C (5), respectively. Configurational dif-

TABLE 2. Melting points of isomeric cyclopentanoid analogs of phosphatidic acid (free acids and esters)

		Melting Point °C					
Com- pound	Con- figuration	Free Acid	Diphenyl Ester	l Dimethyl Ester			
4	1,2,3/0-(1P)	76-78	52-53	34-36			
5	1,2/3-(3P)	77.5-78.5	56 - 57	53 - 54			
6	1,2/3-(1P)	78-80	43-44	68 - 71			
7	1,3/2-(1P)	82 - 83.5	57 - 58	59 - 60			
8	1,3/2-(2P)	73-74.5	69-70	61-63			

ferences among the analogs have a greater effect on the melting points of the dimethyl and diphenyl esters than on the melting points of the free acids themselves (Table 2). Similarly, the chromatographic behavior of the esters is more affected than is that of the free acids (see below). The 1,2,3/0 esters, with one exception, are lower melting than the corresponding members of the other two series, a characteristic observed also for the 1,2,3/0 benzyl ether 35, and noted previously for the 1,2,3/0 trishomoacylcyclopentanetriols (2).

Spectroscopic analysis

(i) Structural and conformational assignments for epoxides 19 and 20 (Scheme 2) were based on earlier observations (14) that in pairs of syn or anti substituted cyclopentane epoxides, the frequency of a strong band in the 830-855 cm⁻¹ region of the IR spectrum is diagnostic. Bands at 854 cm⁻¹ for 19 and 843 cm⁻¹ for 20 established the configurations. This was confirmed by NMR (14) by measurement of the coupling of the cyclopentanoid ring proton on C-1 with the vicinal oxirane proton of C-2. For 19 and 20 this coupling was 1.2 Hz and 0.6 Hz, respectively, in agreement with the assignment. In both compounds, the conformation, as determined from the coupling of the cyclopentanoid ring proton on C-1 with the vicinal methylenic protons on C-5 (14), showed the expected endo, or boat, conformation. The chemical shifts of the benzylic CH₂ resonances in 19 and 20 differed, respectively, by 0.19 ppm (C₆D₆) and 0.08 ppm (CDCl₃). Similar differences were observed in the bis-p-nitrobenzoates 21b and 22b (0.14 ppm in CDCl₃). These differences permitted the determination of the isomeric composition of mixtures and the isomeric purity of each of the four compounds after separation.

(*ii*) O-Isopropylidene monobenzyl ethers 14 and 33. The NMR spectra were consistent with the proposed structures. The gem-dimethyldioxolane moieties in both compounds gave signals for *exo*- and *endo*-CH₃ groups within the ranges reported earlier (22).

(iii) Di-O-palmitoyl monobenzyl ethers 16, 23, 26,

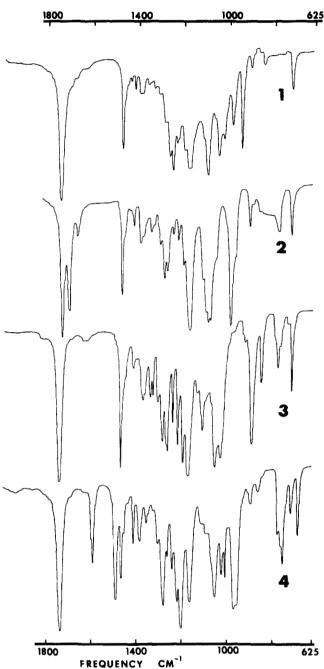


Fig. 2. Infrared spectra of KBr dispersions of the (1,3/2)-2P analog 8 and some of its derivatives. Spectrum 1 is of the free acid; 2 is of the dipotassium salt trihydrate (see text and Table 1); 3 is of the dimethyl ester; 4 is of the diphenyl ester.

and 35. In the spectra of 26 and 35, the nonequivalence (23, 24) of the benzylic CH_2 protons is just enough to produce a narrow doublet representing the inner lines of the expected AB quartet (25). 100 MHz spectra show the AB quartet clearly.

(*iv*) PA analogs and PA dimethyl and diphenyl esters. The NMR spectra of 4-8 showed the high field resonance signals expected for the aliphatic

chains: δ 0.89, CH₃; δ 1.26, CH₂. Signals for ring CH_2 and ring O-C-H protons were not resolved, the spectra showing only broad lines. Much of the line broadening may be due to phosphate chelated paramagnetic ions (26), probably derived from the platinum catalyst used for cleavage of the phenyl groups. The spectra of the corresponding methyl and phenyl diesters of PA are significantly sharper, and the signals of some of the ring O-C -H protons are resolved. Both 28 and the corresponding dimethyl ester give a signal representing the ring proton (C-2)-H, as a well-resolved triplet, $J_{average} = 4.0$ Hz. This coupling shows that the conformational equilibrium of the ring backbone of this lipid analog is identical with that deduced in the earlier work (27) for the tribenzoate of 3. From this we infer that conformational preference deduced for the simpler compounds is a reasonable basis for predicting the conformation of ring skeletons of the phospholipid analogs.

The NMR spectra of the dimethyl esters of 4, 5, and 6 showed fine structure in the signals of the P-O-CH₃ protons centered at δ 3.80. In addition to the ³¹P-O-C-H coupling of 11.3 Hz there was a further splitting of 0.5-1 Hz, similar to that observed in another system (28, 29). The multiplicity in the signal reflects the chemical nonequivalence of the diastereotopic methyl groups in the PA analogs (30). In the earlier case (23, 24) the prochiral phosphorus atom (23, 31) gave rise to diastereomeric methyl esters.

Infrared spectroscopy

The infrared spectra of KBr dispersions of the PA analogs, their salts, and their dimethyl and diphenyl esters show well-resolved absorption bands (Fig. 2). In general, these spectra agree with what is expected for such compounds (32). Details of these spectra will be discussed in a future publication.

Thin-layer chromatography

The chromatographic mobility of the PA analogs was, in general, slightly greater in acid or neutral solvent systems than that of dipalmitoyl glycerophosphoric acid; in an alkaline system the analogs, in common with glycero-PA, did not migrate (**Table 3**). The dependence of mobility on the ring configuration of the isomers was marginally evident in the chromatography of the free acids, but more marked for the dimethyl and diphenyl esters, where the (1,2,3/0)isomers (e.g., 37) showed greater apparent polarity (Table 3). The PA analogs gave well-defined spots on chromatography in acid or neutral solvent systems, unlike other PA analogs such as the diether SBMB

		R _f						
			Free Acie	1	Diphenyl Esther	Dimethyl Esther		
Com- pound			Solvent		Solvent D	Solvent		
	Configuration	A	В	C		E		
4	1,2,3/0-(1P)	0.36	0.62	0.00	0.40	0.29		
5	1,2/3-(3P)	0.39	0.67	0.00	0.44	0.41		
6	1,2/3-(1P)	0.44	0.70	0.00	0.40	0.39		
7	1,3/2-(1P)	0.44	0.70	0.00	0.46	0.41		
8	1,3/2-(2P)	0.46	0.71	0.00	0.54	0.48		
	Dipalmitoylglycero- phosphoric acid	0.36	0.62	0.00				

Solvent systems: A, $CHCl_3-MeOH-H_2O$ 65:35:5; B, $CHCl_3-(CH_3)_2CO-MeOH-HOAc-H_2O$ 6:8:2:2:1; C, $CHCl_3-MeOH-30\%NH_4OH$ 65:35:5; D, $CCl_4-EtOAc$ 8:2; E, $CHCl_3-Et_2O$ 3:1.

glycero-analogs reported by Kates et al. (5), for which these systems were less suitable.

Rationale

The synthesis of the five PA analogs provides us with reasonable routes to the synthesis of more complex cyclopentanoid lipids. Our ultimate goal is to study the biological behavior of these lipid analogs, and of compounds derived from them. It is clear that, as with other lipid systems (33, 34), physical properties such as the gel-to-liquid crystal transition temperatures should be determined before such studies are undertaken. With this provision, we believe that study of the physical and biological properties of these phospholipid analogs can provide kinds of insight into the molecular organization of lipid and lipoprotein assemblies that are not available with the existing lipid repertoire.

EXPERIMENTAL SECTION

DL-2-Cyclopenten-1-ol⁶ 11

A solution of 450 g NaHCO₃ in 21 of water in a 5-l, three-neck flask was chilled to 0°C and 140 g of freshly prepared DL-3-chloro-1-cyclopentene (2, 35) was added over a period of 1 hr. The reaction mixture was then stirred for an additional 2 hr at 0°C, the product was salted out with solid NaCl (500 g) and recovered by extraction with ether. The extract was dried, the solvent was evaporated, and the oil distilled bp 62–65°C/36 mm; n_D²⁸ 1.4692; R_f 0.30 in CHCl₃–Et₂O 20:1. The TLC showed traces of

dicyclopentenyl ether (R_f 0.75) in all fractions of the distillate, but the impurity did not interfere in subsequent steps. IR spectra were measured on solutions in CS₂. The ether side-product was purified by TLC. IR (CS₂) showed strong bands at 1160, 1315, and 1360 cm⁻¹, none of which were given by the allylic alcohol. The alcohol gave bands at 965 and 775 cm⁻¹ which were weak or absent in the spectrum of the ether.

1-O-Benzyl-2-cyclopentenol 12

A solution of 2-cyclopenten-1-ol 11 (21 g, 25 mmoles) and benzyl chloride (68.3 g, 39 mmoles) in benzene (100 ml) was refluxed with powdered KOH (18 g) in an apparatus equipped with a Dean-Stark phase separator until 5.0 ml of H₂O were collected. The mixture was poured over ice (200 g), the aqueous phase was extracted with CHCl₃, and the extracts were combined with the benzene phase. The solution was washed (ice-cold 1 M H₂SO₄, water, 5% NaHCO₃, water), dried, and evaporated. Excess benzyl chloride was removed by distillation at 30°C (50-60 mm) and 12 was distilled (bp 72-75°C/0.6 mm); (27.2 g; 15.6 mmoles, 62.5%). Spectroscopic data were consistent with the proposed structure, but acceptable elemental analyses could not be obtained. Similar difficulties in the case of an O-isopropylidene cyclopentenediol were reported by Young, Hall, and Winstein (36).

NMR: δ 7.30 [5], s, aromatic; δ 5.95 [2], m, olefinic; δ 4.68 [1], m, ring O-C-H; δ 4.54 [2], s, benzyl CH₂; δ 1.65-2.60 [4], m, ring CH₂.

Benzyl ethers of triols

DL-(1,2/3)-1,2-O-Isopropylidene-3-O-benzylcyclopentane-1,2,3-triol 14. Isopropylidenetriol 13 (3.0 g, 19.0

⁶ We thank Dr. Henry C. Stevens for the details of this procedure.

mmoles) and benzyl chloride (4.8 g, 38 mmoles) were reacted in refluxing benzene in the presence of powdered KOH (3.0g) for 8 hr. Benzyl ether 14 was isolated as described for 12 (3.60 g, 14.5 mmoles, 77%; bp $126-129^{\circ}C/0.5$ mm, mp $15-16^{\circ}C$).

Anal. Calc. for $C_{15}H_{20}O_3$ (248.3): C, 72.55; H, 8.12. Found: C, 72.46; H, 8.29.

NMR: δ 1.28, 1.39 [6], d, endo and exo-CH₃ (22); δ 1.85 [4], s, ring CH₂; δ 4.50 [2], s, benzyl CH₂; δ 7.30 [5], s, C₆H₅; δ 3.87 [1], s (broad), CH-O-CH₂C₆H₅; δ 4.42-4.82 [2], m, ring CH-OIp.

IR (CS₂): 3030, 3060, 3090, 735, 695 cm⁻¹ (aromatic); 1360, 1370, 1375 cm⁻¹ (gem-dimethyl).

DL-(1,2,3/0)-1-O-benzyl-2,3-O-isopropylidenecyclopentane-1,2,3-triol 33. Isopropylidenetriol 32 (2) (12.0 g, 75.9 mmoles) was benzylated in refluxing benzene (120 ml) with benzyl chloride (20 g; 158 mmoles) and KOH (14 g) and the benzyl ether was isolated as described for 12. The product 33 (16.9 g, 68.1 mmoles, 90%, bp 122-125°C/0.4 mm) had GLC retention 1.10 relative to 14.

Anal. Calc. for C₁₅H₂₀O₃ (248.3): C, 72.55; H, 8.12. Found: C, 72.82; H, 8.50.

NMR: δ 1.32, 1.53 [6], d, endo- and exo-CH₃, (22); δ 4.66 [2], s, benzyl CH₂; δ 1.55–2.08 [4], m, ring CH₂; δ 7.33 [5], s, C₆H₅; δ 4.45–4.60 [2], m, ring CH–O–Ip; δ 3.36–3.75 [1], m (unresolved), ring CH-O-CH₂C₆H₅.

IR (liquid film): 3030, 3060, 3090, 1495, 735, 700 cm⁻¹ (aromatic); 1365, 1377 (doublet, shoulder at 1360) cm⁻¹, gem-dimethyl.

(1,2/3)-3-O-Benzylcylopentane-1,2,3-triol 15. METHOD I. Isopropylidenebenzyltriol 14 (2.36 g, 950 mmoles) was hydrolyzed by stirring for 2 hr in refluxing 0.1 N H₂SO₄ (200 ml). The cooled mixture was stirred with 10 g BaCO₃ for 1 hr, filtered through Celite and evaporated (1.65 g, 7.93 mmoles, 84%). TLC (CHCl₃– MeOH–H₂O, 90:10:1) showed one spot, R_f 0.50, GLC showed the presence of 1% of 34, and 1% unindentified impurities. Preparative TLC on silica (CHCl₃– MeOH–H₂O 90:10:1) gave an analytical sample.

Anal. Calc. for $C_{12}H_{16}O_3$ (208.3): C, 69.21; H, 7.75. Found: C, 68.98; H, 7.87.

NMR: δ 7.30 [5], s, C₆H₅, δ 4.54 [2], s, benzyl CH₂; δ 3.10 [2], s (broad), OH; δ 1.30–2.40 [4], m, ring CH₂; δ 3.80–4.20 [3], m, ring O–C–H.

METHOD II. A solution of 1-O-benzylcyclopentenol 12 (30 g; 0.17 moles) in 900 ml of EtOH-dioxane-H₂O 1:1:1 containing NaOH (12.5 g; 0.31 mole) was maintained at -20° C while a cool solution of KMnO₄ (19 g; 0.2 moles) in water (700 ml) was added (4 hr). After a further hour, MnO₂ was reduced with SO₂ gas. The solution was filtered, concentrated, and continuously extracted with CH₂Cl₂ (2 days) to give a yellow oil (21.0 g, 0.11 mole, 64%) that cochromatographed with 15 as prepared above. The product was purified by conversion to 14, which was then distilled; the fraction bp $95-100^{\circ}$ C/0.5 mm (16.5 g) was >96% pure (GLC) and free from 33. The *bis-p*-nitrobenzoate derivative (mp 117-118°C) was analyzed.

Anal. Calc. for $C_{26}H_{22}O_9N_2$ (506.5): C, 61.66; H, 4.38; N, 5.53. Found: C, 61.70; H, 4.32; N, 5.31.

(1,2,3/0)-1-O-Benzylcyclopentane-1,2,3-triol 34. Compound 33 (6.2 g, 25 mmoles) was hydrolyzed in refluxing 0.1 N H₂SO₄ (300 ml) for 3 hr. Workup as described for 15 gave 34 in quantitative yield. TLC (CHCl₃-MeOH-H₂O 90:10:1) showed one spot (R_f 0.60); 97% pure by GLC. The bis-p-nitrobenzoate derivative melted at 107-109°C.

Anal. Calc. for $C_{26}H_{22}O_9N_2$ (506.5): C, 61.66; H, 4.38; N, 5.53. Found: C, 61.73; H, 4.38; N, 5.61. NMR: δ 7.30 [5], s, C_6H_5 ; δ 4.52 [2], s, benzyl CH₂; δ 1.80 [4], s, ring CH₂; δ 3.17 [2], s, OH; δ 3.85 [3],

ring O-C-H.

DL-(1,2,3/0)-1-O-Benzyl-2,3-epoxy-1-cyclopentanol 19 and DL-(1/2,3)-1-O-benzyl-2,3-epoxy-1-cyclopentanol 20. A solution of *m*-chloroperoxybenzoic acid (19 g; 110 mmoles) in CHCl₃ (300 ml) was cooled to 3-5°C and stirred while DL-1-O-benzyl-2-cyclopenten-1-ol 12 (15 g; 86 mmoles) in 15 ml of CHCl₃ was added during 30 min. Four such reactions were carried out simultaneously. After standing overnight in the dark at 5°C, the suspensions were filtered, combined, concentrated, and the residue was extracted with cool benzene (100 ml). The extract was filtered and washed with 5% NaHCO₃ and water, dried, and evaporated. Distillation (117-120°C/0.5 mm) gave an oil (45 g, 24 mmoles; 69%) containing two epoxides 20 and 19. A portion was separated by preparative TLC. R_f (CHCl₃-Et₂O 20:1): 20, 0.70; 19, 0.80. Ratio anti/syn 2:1 by GLC.

Anal. Calc. for C₁₂H₁₄O₂ (190.2): C, 75.76; H, 7.42. Found: C, 75.84; H, 7.58.

(1,3/2)-1-O-Benzylcyclopentane, 1,2,3-triol 22a and (1,2/3)-1-O-benzyl-cyclopentane-1,2,3-triol 21a. The mixture of 19 and 20 (25 g, 0.19 moles) was suspended in 500 ml of 0.1 N H₂SO₄ and stirred at reflux temperature (3 hr). The solution was neutralized with BaCO₃ (15 g), filtered, and continuously extracted with CH₂Cl₂ (24 hr). The extract was concentrated to an oil (19.8 g, 73%). TLC (CHCl₃-MeOH-H₂O 90:10:1) showed two major components (R_f 0.40 and 0.55). GLC (trimethylsilyl derivatives) showed that 21a and 22a were formed in a ratio of 35:65.

DL-(1,2/3)-1-O-Benzyl-2,3-bis-O-(p-nitrobenzoyl)-cyclopentane-1,2,3-triol 21b and DL-(1,3/2)-1-O-benzyl-2,3-bis-O-(p-nitrobenzoyl)-cyclopentane-1,2,3-triol 22b. p-Nitrobenzoyl chloride (55 g, 0.30 moles) was added to a solution of 21a and 22a (20 g, 0.13 moles) in pyridine

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(50 ml), chilled to 0°C and the mixture was left overnight. Ice (100 g) was added and after 1 hr the slurry was diluted with water and extracted with CHCl₃. The CHCl₃ solution, worked up as usual, gave a solid product (55 g). TLC (hexane-Et₂O 60:40) showed a single spot $(R_f 0.40)$ containing 21b and 22b, and a minor spot $(R_f \ 0.35)$ (*p*-nitrobenzoic anhydride). The isomers were separated by fractional recrystallization from EtOAc; the rate of crystallization was adjusted for optimum separation (several crops of approximately 2 g each were collected over a period of a week). Isomeric distribution was assayed by integration of the NMR signals at $\delta 4.5 - 4.6$ (benzylic CH₂). Early crops were almost pure 21b (mp 159-162°C) while later crops were enriched in 22b. Subsequent recrystallization of the later crops from acetonitrile gave pure 22b (mp 122-123°C).

Anal. Calc. for $C_{26}H_{22}O_9N_2$ (506.5): C, 61.66; H, 4.38; N, 5.53. Found: 22b: C, 61.76; H, 4.15; N, 5.50. Found 21b: C, 61.76; H, 4.48; N, 5.48.

NMR 22b: δ 4.6, benzyl CH₂; δ 7.3, C₆H₅; 21b: δ 4.5, benzyl CH₂; δ 7.2, C₆H₅.

DL-(1,3/2)-1-O-Benzylcyclopentane-1,2,3-triol 22a or DL-(1,2/3)-1-O-benzylcyclopentane, 1,2,3-triol 21a. The appropriate bis-p-nitrobenzoate 21b or 22b was refluxed for 2 hr in 250 ml of 10% KOH in MeOH– H₂O 40:60. After cooling and neutralization with 6 N H₂SO₄, K₂SO₄ was removed; the filtrate was concentrated and then extracted continuously with CH₂Cl₂ for 24 hr. The extract, dried and concentrated to an oil, gave yields of 95–99%. Each diol was isomerically pure and was used at once for the next stage of the synthesis.

O-Benzyl-di-O-palmitoyl triols

DL-(1,2/3)-3-O-Benzyl-1,2-di-O-palmitoylcyclopentane-1,2,3-triol 16. Benzyl diol 15 (3.33 g, 16 mmoles) in 20 ml of pyridine at 0°C was acylated with palmitoyl chloride (10 g, 36 mmoles) (2). The oily product 16 crystallized on treatment with ethanol at 5°C (10.9 g, 15.9 mmoles, 99%). TLC (hexane-Et₂O-HOAc 90:10:1) showed a major component, R_f 0.23. Trituration with cold acetone removed the color; traces of fatty acid were removed by recyrstallization at 4°C (1 g in 30 ml CH₃OH); mp 52–53°C).

Anal. Calc. for C₄₄H₇₆O₅ (685.1): C, 77.14; H, 11.18. Found: C, 77.27; H, 11.26.

NMR: δ 4.56 [2], s, benzylic CH₂.

DL-(1,3/2)-1-O-Benzyl-2,3-di-O-palmitoylcyclopentane-1,2,3-triol 26. Benzyl diol 22a (2.38 g, 11.9 mmoles) was acylated in 25 ml of pyridine with palmitoyl chloride (5.0 g, 18 mmoles) (2) to give 6.30 g of an oil that could not be induced to crystallize. TLC showed a relatively polar impurity and traces of fatty acid. Purification by chromatography on silica gel in hexane-benzene 1:1 gave an oil that crystallized in vacuo mp. 42–43°C (3.54g, 5.17 mmoles, 44%).

Anal. Calc. for C₄₄H₇₆O₅ (685.1): C, 77.14; H, 11.18. Found: C, 77.10; H, 11.09.

NMR: δ 4.58, 4.60 [2], "doublet", benzylic CH₂.

DL-(1,2/3)-1-O-Benzyl-2,3-di-O-palmitoylcyclopentane-1,2,3-triol 23. Benzyl diol 21a (1.38 g, 6,63 mmoles) in 10 ml of pyridine was acylated with palmitoyl chloride (5.0 g, 18 mmoles) (2) to give 4.0 g of an oil, which was then purified by silica chromatography in hexane-benzene 1:1 giving 23 (3.67 g, 5.36 mmoles, 81%) mp 44-45°C.

Anal. Calc. for C₄₄H₇₆O₅ (685.1): C, 77.14; H, 11.18. Found: C, 77.22; H, 10.94.

NMR: δ 4.50 [2], s, benzylic CH₂.

DL-(1,2,3/0)-1-O-Benzyl-2,3-di-O-palmitoylcyclopentane-1,2,3-triol 35. Diol 34 (3.33 g, 16 mmoles) in 20 ml of pyridine was acylated with palmitoyl chloride (10 g, 36 mmoles) (2). Purification by column chromatography gave 35 (7.18 g, 10.5 mmoles, 66%) which crystallized slowly in vacuo and was recrystallized from MeOH at 4°C (mp. 38–39°C).

Anal. Calc. for C₄₄H₇₆O₅ (685.1): C, 77.14; H, 11.18. Found: C, 77.16; H, 11.32.

NMR: δ 4.50, 4.55 [2] "doublet", benzylic CH₂.

Di-O-palmitoyl triols

DL-(1,2/3)-1,2-Di-O-palmitoylcyclopentane-1,2,3-triol 17. Diacylmonobenzyl triol 16 (8.0 g; 11.7 mmoles) in 90 ml of CHCl₃-MeOH 75:15 was vigorously stirred under hydrogen with 2 g of 10% Pd-C. After 2 hr the catalyst was removed by filtration over Celite; evaporation gave 17 in quantitative yield; recrystallization (hexane) gave an analytical sample mp 63-64°C.

Anal. Calc. for $C_{37}H_{70}O_5$ (594.9): C, 74.69; H, 11.86. Found: C, 74.51; H, 11.89.

DL-(1,2,3/0)-1,2-Di-O-palmitoylcyclopentane-1,2,3-triol 36. Diacylmonobenzyl triol 35 (4.62 g, 6.75 mmoles) in 20 ml of EtOAc was hydrogenolyzed as above (1.5 g Pd-C; 90 min). Recrystallization from hexane gave 36 (3.98 g, 99%; mp 64–65°C).

Anal. Calc. for C₃₇H₇₀O₅ (594.9): C, 74.69; H, 11.86. Found: C, 74.61; H, 11.74.

DL-(1,2/3)-2,3-Di-O-palmitoylcyclopentane-1,2,3-triol 24. Compound 23 (3.67 g; 5.36 mmoles) in 15 ml of EtOAc was hydrogenolyzed (1 g Pd-C, 2.5 hr). TLC (CHCl₃-Et₂O 20:1) on sodium acetate-impregnated plates gave one spot (R_f 0.55), but unbuffered silica caused acyl migration as evidenced by a second spot (R_f 0.65). The product 24 (3.11 g, 5.23 mmole; 98%) was recrystallized from hexane (mp 52-53°C).

Anal. Calc. for C₃₇H₇₀O₅ (594.9): C, 74.69; H, 11.86. Found: C, 74.59; H, 11.66. DL-(1,3/2)-1,2-Di-O-palmitoylcyclopentane-1,2,3-triol 27. Compound 26 (3.12 g, 4.55 mmoles) in 20 ml of EtOAc was hydrogenolyzed (1 g Pd-C) for 2 hr. The product crystallized during evaporation (2.70 g, 4.56 mmoles, 100%). Recrystallization (hexane) gave 27, mp 57-59°C.

Anal. Calc. for $C_{37}H_{70}O_5$ (594.9): C, 74.69; H, 11.86. Found: C, 74.65; H, 11.72.

Diphenyl esters of phosphatidic acids

DL-(1,2/3)-3-O-(Diphenylphosphoryl)-1,2-di-O-palmitoylcyclopentane-1,2,3-triol 18. Diacyl triol 17 (6.0 g, 11 mmoles) in 1:1 anhydrous pyridine–Et₂O (25 ml) was stirred at 0°C while DPPC (5.37 g, 20 mmoles) was added; stirring was continued overnight at room temperature. Ice (2 g) was added and after 30 min the mixture was diluted with 100 ml each of CHCl₃ and water. The CHCl₃ phase was washed with water, dried, and evaporated to an oil that crystallized upon addition of EtOH (7.64 g, 9.24 mmoles; 84%); TLC (CCl₄-EtOAc 80:20) showed one major spot, R_f 0.44; recrystallization from MeOH gave 18, mp 56–57°C.

Anal. Calc. for $C_{49}H_{79}O_8P(827.1)$: C, 71.15; H, 9.63; P, 3.75. Found: C, 71.09; H, 9.50; P, 3.76.

DL-(1,2/3)-1-O-(Diphenylphosphoryl)-2,3-di-O-palmitoylcyclopentane-1,2,3-triol 25. Diacyl triol 24 (2.77 g, 4.66 mmoles) in 20 ml of pyridine was phosphorylated (DPPC 2.70 g, 10 mmoles) as described for 18. The oily product, which crystallized slowly (3.62 g; 6.09 mmoles, 94%), was chromatographically pure, R_f 0.40 (CCl₄-EtOAc 80:20). Recrystallization from MeOH gave 25, mp 43-44°C.

Anal. Calc. for C₄₉H₇₉O₈P (827.1): C, 71.15; H, 9.63; P, 3.75. Found: C, 71.33; H, 9.86; P, 3.72.

DL-(1,2,3/0)1-O-(Diphenylphosphoryl)-2,3-di-O-palmitoylcyclopentane-1,2,3-triol 37. Diacyl triol 36 (3.60 g, 6.05 mmoles) in 25 ml of pyridine was phosphorylated (DPPC 3.80 g, 14.1 mmoles) as described for 18, giving chromatographically pure 37 (4.80 g, 5.81 mmoles; 96%); R_f 0.36 (CCl₄-EtOAc 80:20). Recrystallization from MeOH gave mp 52–53°C.

Anal. Calc. for C₄₉H₇₉O₈P (827.1): C, 71.15; H, 9.63; P, 3.75. Found: C, 71.43; H, 9.68; P, 3.73.

DL-(1/2,3)-1-O-(diphenylphosphoryl)-2,3-Epoxy-1-cyclopentanol 30. To epoxyalcohol 29 (2) (3.0 g; 30 mmoles) in 50 ml of pyridine at 0°C, DPPC (10.8 g; 40 mmoles) in 15 ml Et₂O was added, and the solution was stirred at room temperature overnight. Ice (10 g) was added, stirred for 1 hr, and 200 ml each of CHCl₃ and water were added. The aqueous phase was extracted with CHCl₃ and the combined CHCl₃ solution was washed with water and dried. Evaporation gave an oil 30 (9.30 g, 28 mmoles; 93%); TLC (CHCl₃-Et₂O 3:1) showed a major phosphate-positive spot, R_f 0.80. An analytical sample was obtained by preparative TLC on silica with $CHCl_3-Et_2O$ 3:1 as developing solvent and $CHCl_3-MeOH-Et_2O$ 1:1:1 as eluant.

Anal. Calc. for $C_{17}H_{17}O_5P$ (332.3): C, 61.45; H, 5.16; P, 9.32. Found: C, 61.63; H, 5.23; P, 9.58.

The slightly impure product was used for hydrolysis to 31.

DL-(1,3/2)-1-O-(Diphenylphosphoryl)-cyclopentane-1,2, 3-triol 31. The phosphorylated epoxide 30 (5.55 g, 16.7 mmoles) in a mixture of 0.4 N H₂SO₄ (40 ml) and dioxane (50 ml), was refluxed for 90 min, cooled, and stirred with 1.4 g NaHCO₃ for 1 hr. Evaporation gave an oily solid, which was triturated with CHCl₃ and filtered. The filtrate was dried and evaporated to give an oil 31 (5.71 g, 16.3 mmoles; 98%); TLC, R_f 0.35 (CHCl₃-MeOH-H₂O 90:10:1). The product was used directly for acylation to 28. For analytical purposes, a portion of 31 was converted to the dibenzoate (14); recrystallization from aqueous EtOH gave needles mp 132–133°C.

Anal. Calc. for C₃₁H₂₇O₈P (558.5): C, 66.66; H, 4.87; P. 5.55. Found: C, 66.57; H, 4.73; P, 5.33.

DL-(1,3/2)-1-O-(Diphenylphosphoryl)-2,3-di-O-palmitoylcyclopentane-1,2,3-triol 28. METHOD 1. Diacyl triol 27 (2.60 g, 4.37 mmoles) in 25 ml of anhydrous pyridine was phosphorylated (DPPC 2.30 g, 8.56 mmoles) and worked up as described for 18. The product (3.39 g, 4.10 mmoles, 94%) was chromatographically pure, R_f 0.46 (CCl₄-EtOAc 80:20). Recrystallization from MeOH gave 28 mp 57.5-58°C. METHOD 2. The phosphorylated triol 31 (1.07 g; 3.05 mmoles) in pyridine (30 ml) was acylated with palmitoyl chloride (2.20 g; 8.0 mmoles), and worked up as described for tris-homoacyl triols (2). TLC (CCl₄-EtOAc 80:20) showed a major spot R_f 0.46. Purification by silicic acid chromatography gave 28, 1.40 g (75%) mp 56-57°C.

Anal. Calc. for $C_{49}H_{79}O_8P$ (827.1): C, 71.15; H, 9.63; P, 3.75. Found: C, 71.25; H, 9.37; P, 3.86.

(1,3/2)-1,3-O-Nitrobenzylidenecyclopentane-1,2,3-triol 38. Triol 3 (15 g, 127 mmoles) in 250 ml of toluene was refluxed for 2.5 hr with *p*-nitrobenzaldehyde (22.6 g, 150 mmoles) and *p*-toluenesulfonic acid (50 mg) in an apparatus fitted with a phase-separating head. The hot solution, decanted from the tarry residue, crystallized (25 g) on cooling, and yielded a second crop (1.5 g) on concentration to 100 ml. The crystals were dissolved in 120 ml toluene at 80–85°C, and the clear solution was decanted from a small quantity of dark oil and allowed to crystallize (14.6 g, 58.1 mmoles, 46%); TLC showed a major spot R_f 0.35 (CHCl₃-Et₂O 3:1). For analytical purposes the TROC ester was prepared: acetal 38 (200 mg; 0.80 mmoles) in Et₂O (5 ml) and pyridine (1 ml) was stirred at 0°C with 2,2,2-trichloroethoxychloroformate (200 mg; 0.94 mmoles) for 16 hr. Ice (2 g) was added; the mixture was stirred for 30 min, and the ester was extracted with Et₂O (340 mg; 0.80 mmoles, 100%). TLC (CHCl₃-Et₂O 20:1) showed one spot, R_f 0.70. Recrystallization from ether gave yellow crystals, mp 143-144°C.

Anal. Calc. for $C_{15}H_{14}O_7NCl_3$ (426.6): C, 42.23; H, 3.31; N, 3.28; Cl, 24.99. Found: C, 42.31; H, 3.44; N, 3.18, Cl, 24.90.

NMR 38: δ 2.15 [4], s, ring CH₂; δ 1.90 [1] broad, OH; δ 4.20 [1] s, (broad) ring HOCH; δ 4.52 [2] s (broad), ring ethereal–O–CH; δ 5.82 [1], s, benzylidene CH; 7.55–8.26 [4] quartet, aromatic. TROC ester of 38: δ 2.23 [4] s, (broad) ring CH₂; δ 5.88 [1], s, benzylidene CH; δ 5.05 [1], ester–O–CH; δ 4.78 [4], COOCH₂CCl₃ and ring ethereal O–C–H.

IR: 38: (CCl₄), 3640 cm⁻¹, OH. (KBr) 3350 cm⁻¹ (broad), OH; 1350, 1525 cm⁻¹, NO₂; 3060, 1610, 750, 700 cm⁻¹, aromatic.

(1,3/2)-1,3-O-(p-Nitrobenzylidene)-2-O-(diphenylphosphoryl)-cyclopentane-1,2,3-triol 39. Acetal 38 (4.0 g; 16.2 mmoles) was phosphorylated in 30 ml of pyridine with DPPC (6.46 g; 24 mmoles), and 39 was isolated as described above. The oil (7.73 g; 16 mmoles, 99%) was decolorized with charcoal in hot benzene; evaporation gave an oil that crystallized in vacuo, mp 83-88°C. TLC (CHCl₃-Et₂O 20:1) showed a major spot R_f 0.60. Recrystallization of a portion (MeOH) removed traces of p-nitrobenzaldehyde and gave yellow plates, 39 mp 88-89°C.

Anal. Calc. for $C_{24}H_{22}O_8PN$ (483.4): C, 59.63; H, 4.59; P, 6.41; N, 2.90. Found: C, 59.96; H, 4.57; P, 6.27; N, 2.80.

NMR: δ 7.55–8.26 [4], quartet, C₆H₄NO₂; δ 7.30 [10], s, C₆H₅; δ 5.82 [1], s, benzylidene CH; δ 4.92 [1], doublet of triplets, ring POC–H; δ 4.70 [2], s, ring (C-1 and C-3)–H; δ 2.10 [4] s, ring CH₂.

(1,3/2)-2-O-(Diphenylphosphoryl)-cyclopentane-1,2,3triol 40. The phosphorylated acetal 39 (2.75 g; 5.69 mmoles) in 50 ml EtOH-CHCl₃ 3:2 was hydrogenolyzed (Pd-C, 4.0 g). Filtration through Celite and concentration gave a yellow solid that was partitioned between CHCl₃ and ice-cold 1 N H₂SO₄. The CHCl₃ phase was washed with 5% NaHCO₃, dried, and evaporated to give 40 as an oily solid (1.75 g, 88%). TLC (CHCl₃-MeOH-H₂O 90:10:1) showed a major spot R_f 0.50. For analytical purposes, a sample was converted to the *bis*-(*p*-nitrobenzoate) derivative: recrystallization (EtOAc) gave yellow needles, mp 195-196°C (partial sublimation at 170°C).

Anal. Calc. for $C_{31}H_{25}O_{12}PN_2$ (648.5): C, 57.41; H, 3.89; N, 4.32, P, 4.78. Found: C, 57.64; H, 3.80; N, 4.31; P, 4.79. The diol discolored on standing and was therefore used at once in the next stage of synthesis.

(1,3/2)-2-O-(Diphenylphosphoryl)-1,3-di-O-palmitoylcyclopentane-1,2,3-triol 41. The phosphorylated triol 40 (2.63 g, 7.50 mmoles) in 20 ml pyridine was acylated with palmitoyl chloride (5.0 g; 18.2 mmoles) at 0°C. Ice was added (10 g) and after 1 hr 41 was isolated as described for the benzyl diacyl derivative 16. The oil crystallized on storage (5.71 g, 6.90 mmoles; 92%). TLC (CCl₄-EtOAc 80:20) showed 41 (R_f 0.54), traces of 40 and palmitic acid. An analytical sample was obtained by preparative TLC; the eluted product was recrystallized from MeOH (mp 69–70°C).

Anal. Calc. for C₄₉H₇₉O₈P (827.1): C, 71.15, H, 9.63; P, 3.75. Found: C, 71.39; H, 9.56; P, 3.61.

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REFERENCES

- IUPAC-IUB Commission of Biochemical Nomenclature. 1968. Tentative rules for the nomenclature of cyclitols. Arch. Biochem. Biophys. 128: 269–279.
- Hancock, A. J., S. M. Greenwald, and H. Z. Sable. 1975. Analogs of natural lipids. I. Synthesis and properties of tris-homoacyl derivatives of cyclopentane-1,2,3-triols. J. Lipid Res. 16: 300-307.
- Dittmer, J. C., and R. L. Lester. 1964. A simple, specific spray for the detection of phospholipids on thin layer chromatograms. J. Lipid Res. 5: 126–127.
- 4. Cubero Robles, E., and D. Van Den Berg. 1969. Synthesis of lecithins by acylation of O-(sn-glycero-3phosphoryl)choline with fatty acid anhydrides. *Biochim. Biophys. Acta.* 187: 520-526.
- Kates, M., C. E. Park, B. Palameta, and C. N. Joo. 1971. Synthesis of diphytanyl ether analogues of phosphatidic acid and cytidine diphosphate diglyceride. *Can. J. Biochem.* 49: 275–281.
- Bligh, E. G., and W. J. Dyer. 1959. A rapid method of total lipid extraction and purification. *Can. J. Biochem. Physiol.* 37: 911-917.
- de Boer, T. J., and H. J. Backer. 1963. Diazomethane. In Org. Syn. Coll. Vol. 4. John Wiley and Sons, New York, N. Y. 250-253.
- Kates, M., and A. J. Hancock. 1971. Determination of ionizable acid groups in phosphatidyl glycerophosphate (diphytanyl ether analogue) by proton magnetic resonance spectroscopy. *Biochim. Biophys. Acta.* 248: 254-262.
- 9. Henbest, H. B., and R. A. L. Wilson. 1957. Aspects of stereochemistry. Part I. Stereospecificity in formation



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of epoxides from cyclic allylic alcohols. J. Chem. Soc.: 1958-1965.

- Franks, J. A., Jr., B. Tolbert, R. Steyn, and H. Z. Sable. 1965. Studies on cyclic polyols. III. The configurations and reactions of some epoxydiols of cyclopentane. *J. Org. Chem.* 30: 1440–1446.
- Buchanan, J. G., and H. Z. Sable. 1972. Stereoselective epoxide cleavages. *In* Selective Organic Transformations, Vol. 2. B. S. Thyagarajan, editor. Wiley-Interscience, New York, N. Y. 1–95.
- 12. Serdarevich, B. 1967. Glyceride isomerization in lipid chemistry. J. Amer. Oil Chem. Soc. 44: 381-393.
- 13. Sable, H. Ž., and T. Posternak. 1962. Recherches dans la série des cyclitols. XXXII. Sur les configurations des cyclopentènediols-3,5 et des cyclopentanediols-1,3. *Helv. Chim. Acta.* **45:** 370-375.
- Steyn, R., and H. Z. Sable. 1969. Studies on cyclitols. XIII. Synthesis and stereochemistry of cyclopentane triols and related epoxycyclanols. *Tetrahedron.* 25: 3579–3597.
- 15. Baer, E., J. Maurukas, and M. Russell. 1952. Synthesis of enantiomeric α-cephalins. J. Amer. Chem. Soc. 74: 152–157.
- 16. Baer, E., and J. Maurukas. 1955. The diazometholysis of glycerophosphatides. A novel method of determining the configuration of phosphatidyl serines and cephalins. J. Biol. Chem. 212: 39-48.
- 17. Augustine, R. L. 1965. Catalytic Hydrogenation. Marcel Dekker, Inc., New York, N. Y. 135-136.
- 18. Wood, H. B., Jr., and H. G. Fletcher, Jr. 1956. Migration of a mesitoyl group from C_1 to C_2 in α -D-glucopyranose. Derivatives of 2-O-mesitoyl-D-glucose. J. Amer. Chem. Soc. **78**: 2849–2851.
- Chapman, D., and D. J. Fluck. 1966. Physical studies of phospholipids. III. Electron microscope studies of some pure fully saturated 2,3-diacyl-pL-phosphatidylethanolamines and phosphatidylcholines. J. Cell Biol. 30: 1-11.
- Williams, R. M., and D. Chapman. 1971. Phospholipids, liquid crystals and cell membranes. Prog. Chem. Fats Other Lipids. 11: 3-79.
- Bangham, A. D., J. deGier, and G. D. Greville. 1967. Osmotic properties and water permeability of phospholipid liquid crystals. *Chem. Phys. Lipids.* 1: 225– 246.
- Sable, H. Z., W. M. Ritchey, and J. E. Nordlander. 1969. Studies on cyclitols. XIV. Methyl group signals of gem-dimethyldioxolanes as indicators of anisotropy. Spectrosc. Lett. 2: 239-245.

- 23. Mislow, K., and M. Raban. 1967. Stereoisomeric relationships of groups in molecules. *In* Topics in Stereochemistry, Vol. 1. N. L. Allinger and E. L. Eliel, editors. Interscience, New York. 1–28.
- 24. Jennings, W. B. 1975. Chemical shift non-equivalence in prochiral groups. *Chem. Rev.* **75:** 307-322.
- Jackman, L. M., and S. Sternhell. 1969. Applications of Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry. Pergamon Press, Braunschweig, Germany. 129–130.
- Jackman, L. M., and S. Sternhell. 1969. Application of Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry. Pergamon Press, Braunschweig, Germany. 9.
- 27. Steyn, R., and H. Z. Sable. 1971. Studies on cyclitols. XVI. Conformational analysis of substituted cyclopentanes, cyclopentenes and cyclopentane oxides. *Tetrahedron.* **27**: 4429-4447.
- Hancock, A. J. 1972. Structural studies on phytanylether membrane lipids in *Halobacterium cutirubrum*. Ph.D. Dissertation. University of Ottawa, Ottawa, Ontario, Canada.
- 29. Hancock, A. J., and M. Kates. 1973. Synthesis of sulfate esters of phosphatidylglycerol (diphytanyl ether analog). *J. Lipid Res.* 14: 430–437.
- Gallo, A. A., A. J. Hancock, and H. Z. Sable. 1977. Analogs of natural lipids. III. Nonequivalence of methyl groups in methylated phospholipids. J. Lipid Res. 18: 77-80.
- 31. Hanson, K. R. 1966. Applications of the sequence rule. I. Naming the paired ligands g,g at a tetrahedral atom X_{ggli} . II. Naming the two faces of a trigonal atom Y_{ghi} . J. Amer. Chem. Soc. 88: 2731–2742.
- Abramson, M. B., W. T. Norton, and R. Katzman. 1965. Study of ionic structures in phospholipids by infrared spectra. J. Biol. Chem. 240: 2389-2395.
- Dennis, E. A. 1973. Phospholipase A₂ activity towards phosphatidylcholine in mixed micelles: surface dilution kinetics and the effect of thermotropic phase transitions. Arch. Biochem. Biophys. 158: 485-493.
- Trauble, H., and H. Eibl. 1974. Electrostatic effects on lipid phase transitions: Membrane structure and ionic environment. *Proc. Nat. Acad. Sci. USA.* 71: 214-219.
- Moffett, R. B. 1963. Cyclopentadiene and 3-chlorocyclopentene. Org. Syn. Coll. 4: 779–785.
- Young, W. G., H. K. Hall, Jr., and S. Winstein. 1959. 1,2- and 1,4-dibromides from cyclopentadiene. J. Amer. Chem. Soc. 78: 4338-4344.